

REVISED ENVIRONMENTAL ASSESSMENT

1. **Date** May 24, 1999

2. Name of Applicant/Petitioner Ticona

3. Address All communications on this matter are to be sent

in care of Counsel for Petitioner, Jerome H. Heckman, Keller and Heckman LLP, 1001 G Street, N.W., Suite 500 West, Washington, D.C.

20001. Telephone: (202) 434-4110.

4. Description of the Proposed Action

The action requested in this Petition is the amendment of an existing Food Additive Regulation to expand the permitted uses of olefin polymers in articles or components of articles that may contact food. Its specific purpose is to amend 21 C.F.R. § 177.1520(a)(3) to provide for use of ethylene-2-norbornene copolymers containing not less than 30 but no more than 70 mole percent of polymer units derived from norbornene. Such copolymers would be limited to use with dry foods containing no free fat or oil (Food Type VIII, identified in § 176.170(c), Table 1).

We note that, while FDA promulgated regulations, effective August 29, 1997, amending its requirements for environmental assessments (EA's), the Agency has not yet made available guidance documents for preparing EA's under the new regulations.

Consequently, the EA presented here has been prepared in accordance with the format requirements that previously appeared at 21 C.F.R. § 25.31a. modified as appropriate to focus on the impact of use and disposal of the subject polymers, in keeping with the new regulations. Since the requirements set forth under new Section 25.40 are less extensive than

000303 EA-Z

98F-0569

the former requirements, we understand that an EA prepared in accordance with the previous formats will be accepted.

The subject ethylene-norbornene copolymers offer several technical properties that make them useful in a variety of food, pharmaceutical, and medical device applications. In particular, the moisture barrier properties of the polymers make them useful in food and pharmaceutical flexible packaging, and in certain rigid packaging applications (e.g., vials and bottles). The polymers also offer good clarity, and a high heat deflection temperature. The latter is of importance in applications involving steam autoclave treatment of the product.

The uses for the polymers at issue in this petition are limited to dry food contact.

These foods are generally packaged either in flexible film or paper and paperboard packaging. They are not generally packaged in rigid containers and are not subject to retort sterilization. Thus, for the applications covered in this petition, the copolymers are expected to be used primarily in film form to produce flexible packaging for dry foods use, where the moisture barrier properties of the polymer will be of greatest value. In addition, the polymers may also be used to produce containers (e.g., bins) that are used to hold bulk quantities of dry food and are generally subject to repeated-use.

The Petitioner does not intend to produce finished food packaging materials from the subject ethylene-norbornene copolymers. Rather, the copolymers will be sold to manufacturers engaged in the production of food-contact materials. Food-contact materials produced with the use of the copolymers will be utilized in patterns corresponding to the national population density and will be widely distributed across the country. Therefore, it is

anticipated that disposal will occur nationwide, with about 76% of the materials being deposited in land disposal sites, and about 24% combusted.¹

The types of environments present at and adjacent to these disposal locations are the same as for the disposal of any other food-contact material in current use. Consequently, there are no special circumstances regarding the environment surrounding either the use or disposal of food-contact materials prepared from ethylene-norbornene copolymers.

5. Identification of Substance that Is the Subject of the Proposed Action

The additives that are the subject of this Petition are copolymers of ethylene and 2-norbornene prepared with not less than 30 but no more than 70 mole percent of polymer units derived from 2-norbornene. As dealt with by the petitioner, the copolymers are marketed under the trade name Topas® and referred to as "Topas® COC [cyclic olefin copolymers]."

The Chemical Abstracts Service (CAS) Registry Number for Topas® COC is 26007-43-2; the CAS name is Bicyclo[2.2.1]hept-2-ene, polymer with ethene. The molecular formula is $(C_7H_{10}\cdot C_2H_4)_x$. The structural formulae for the polymer repeating units are as follows:

⁻ Characterization of Municipal Solid Waste in the United States: 1997 Update, EPA 530-R-98-007, U.S. Environmental Protection Agency (5305W), Washington DC, 20460, May 1998.

The molecular structure of the polymer consists of varied sequences of the units shown above randomly incorporated along the polymer chain.

6. Introduction of Substances into the Environment

Under 21 C.F.R. § 25.40(a), an environmental assessment ordinarily should focus on relevant environmental issues relating to the use and disposal from use, rather than the production, of FDA-regulated articles. Moreover, information available to the Petitioner does not suggest that there are any extraordinary circumstances in this case indicative of any adverse environmental impact as a result of the manufacture of Topas® COC copolymers. Consequently, information on the manufacturing site and compliance with relevant emissions requirements is not provided here.

No environmental release is expected upon the use of the subject copolymers to fabricate packaging materials for dry foods. In these applications, the polymers are expected largely to be used in film form and will be entirely incorporated into the finished food package. Any waste materials generated in this process, e.g., plant scraps, are expected to be disposed of as part of the packaging manufacturer's overall nonhazardous solid waste in accordance with established procedures.

Disposal by the ultimate consumer of food-contact materials produced by the subject copolymers will be by conventional rubbish disposal and, hence, primarily by sanitary landfill or incineration. The subject olefin copolymers consist of carbon and hydrogen. Thus, no toxic combustion products are expected as a result of the proper incineration of the copolymers.

Only extremely small amounts, if any, of Topas® COC copolymer constituents are expected to enter the environment as a result of the landfill disposal of food-contact articles, in light of the Environmental Protection Agency's (EPA) regulations governing municipal solid waste landfills. EPA's regulations require new municipal solid-waste landfill units and lateral expansions of existing units to have composite liners and leachate collection systems to prevent leachate from entering ground and surface water, and to have ground-water monitoring systems. 40 C.F.R. Part 258. Although owners and operators of existing active municipal solid waste landfills that were constructed before October 9, 1993 are not required to retrofit liners and leachate collections systems, they are required to monitor groundwater and to take corrective action as appropriate. The lack of any leaching is especially true considering that the subject substances are high molecular weight polymers that contain only minute levels of extractable material even under conditions that greatly exaggerate environmental exposure conditions.²

7. Fate of Emitted Substances in the Environment

(a) Air

No significant effect on the concentrations of and exposures to any substances in the atmosphere are anticipated due to the proposed use of Topas® COC copolymers. The

This expectation is confirmed by the results of extraction studies described in the Appendix to this Environmental Assessment. As shown there, when 157-mil thick bars of the polymer were exposed to water, 3% acetic acid, and 15% ethanol at 100°C for two hours, less than 3 ppb of norbornene monomer was found in the extracts. Moreover, due to the very limited food-contact use requested in this Petition (use with dry foods only), the quantity of ethylene-norbornene copolymer in solid waste deposited in landfills will be extremely small.

polymers are of high molecular weight and do not volatilize. Thus, no significant quantities of any substances will be released upon the use and disposal of food-contact articles manufactured with these copolymers.

The products of complete combustion of the polymer would be carbon dioxide and water; the concentrations of these substances in the environment will not be significantly altered by the proper incineration of the polymers in the amounts utilized for food packaging applications.

(b) Water

No significant effects on the concentrations of and exposures to any substances in fresh water, estuarine, or marine ecosystems are anticipated due to the proposed use of the subject copolymers. No significant quantities of any substance will be added to these water systems upon the proper incineration of the polymers, nor upon its disposal in landfills due to the extremely low levels of aqueous migration of polymer components.

(c) Land

Considering the factors discussed above, no significant effects on the concentrations of and exposures to any substances in terrestrial ecosystems are anticipated as a result of the proposed use of the subject ethylene-norbornene copolymers. In particular, the extremely low levels of migration of monomer, even at 100°C, demonstrated by the extraction studies, indicate that virtually no leaching of these substances may be expected to occur under normal environmental conditions when finished food-contact materials are disposed of. Furthermore, the very low production of Topas® COC copolymers for use in food-contact applications precludes any substantial release to the environment of their components. Thus, there is no

expectation of any meaningful exposure of terrestrial organisms to these substances as a result of the proposed use of the copolymers.

Considering the foregoing, we respectfully submit that there is no reasonable expectation of a significant impact on the concentration of any substance in the environment due to the proposed use of Topas® COC copolymers in the manufacture of articles intended for use in contact with dry food.

8. Environmental Effects of Released Substances

As discussed previously, the only substances that may be expected to be released to the environment upon the use and disposal of food packaging materials fabricated with the use of the subject copolymers consist of extremely small quantities of combustion products and extractables. As discussed in Section E of the Petition, two copolymers, having different percentages of polymer units derived from 2-norbornene, were subjected to acute oral toxicity study in rats demonstrating that the LD₅₀ for each test material is greater than 2000 mg/kg bw. In addition, the monomer 2-norbornene is of very low acute toxicity, as demonstrated by an LD₅₀ in excess of 11000 mg/kg bw; and the monomer has been shown to be non genotoxic. Based on these considerations, no adverse effect on organisms in the environment is expected as a result of the disposal of articles containing the copolymers. In addition, the use and disposal of the copolymers are not expected to threaten a violation of applicable laws and regulations, e.g., the Environmental Protection Agency's regulations in 40 C.F.R. part 60 that pertain to municipal solid waste combustors and part 258 that pertain to landfills.

9. Use of Resources and Energy

As is the case with other food packaging materials, the production, use and disposal of Topas® COC copolymers involves the use of natural resources such as petroleum products, coal, and the like. However, the use of the subject copolymers in the fabrication of food-contact materials is not expected to result in a net increase in the use of energy and resources, since the copolymers are intended to be used in place of similar polymers now on the market for use in food packaging applications. Specifically, as discussed in Item 4 above, the subject copolymers are proposed for use in contact with dry foods only. Due to this limitation, the polymers are expected to be used largely in flexible film packaging to serve as a moisture barrier. Dry foods currently are packaged primarily in similar sorts of flexible film or in paper and paperboard packaging. Polymers currently used in such applications include other olefin polymers that are cleared under 21 C.F.R. § 177.1520 as well as other polymers that are used for their moisture barrier properties, such as polyvinylidene chloride (PVDC).

The replacement of these types of materials by Topas® is not expected to have any adverse impact on the use of energy and resources. Manufacture of the copolymers and conversion to finished food packaging materials will consume energy and resources in amounts comparable to the manufacture and use of other polyolefins. Moreover, the film and paper materials currently in use for dry foods packaging are not recovered for recycling to a significant extent but are disposed of by means of sanitary landfill and incineration.

Packaging materials produced from ethylene-norbornene copolymers are expected to be

disposed of according to the same patterns when they are used in place of the current materials. Thus, there will be no impact on current or future recycling programs.

As also noted in Item 4, the subject copolymers may also be used to produce rigid containers for holding food. In this case, based on consideration of current packaging patterns for dry foods, it is expected that the polymers will be limited to use in articles such as bins that are intended for use in holding bulk quantities of dry food. These articles will generally be subject to repeated use and will have a reasonably long service life. Currently used materials that may be replaced by ethylene-norbornene copolymers in these applications are expected to consist of other polymers, including other olefin polymers, acrylics, etc. At the end of their service lives, the articles are expected to be disposed of by standard means (primarily landfill); disposal patterns are expected to be the same as those for the currently used materials. Thus, again there will be no adverse impact on recycling or on solid waste production.

10. Mitigation Measures

As shown above, no significant adverse environmental impacts are expected to result from the use and disposal of food-contact materials fabricated from the subject copolymers. This is primarily due to the minute levels of leaching of potential migrants from the finished article; the insignificant impact on environmental concentrations of combustion products of the polymers; and the close similarity of the subject copolymers to the materials they are intended to replace. Thus, the use of the copolymers as proposed is not reasonably expected to result in any new environmental problem requiring mitigation measures of any kind.

11. Alternatives to the Proposed Action

No potential adverse environmental effects are identified herein which would necessitate alternative actions to that proposed in this Petition. The alternative of not approving the action proposed herein would simply result in the continued use of the materials which the subject copolymers would otherwise replace; such action would have no environmental impact. In view of the excellent qualities of the Topas® COC polymers for use in food-contact applications, the fact that the copolymer constituents are not expected to enter the environment in more than minute quantities upon the use and disposal of finished food-contact articles, and the absence of any significant environmental impact which would result from their use, the promulgation of a Food Additive Regulation to permit the use of Topas® COC polymers as described herein is environmentally safe in every respect.

12. List of Preparers

- a. Michael T. Flood, Staff Scientist, Keller and Heckman LLP, 1001 G Street, N.W., Suite 500 West, Washington, D.C. 20001.
- b. Holly H. Foley, Staff Scientist, Keller and Heckman LLP, 1001 G Street, N.W., Suite 500 West, Washington, D.C. 20001.

13. Certification

The undersigned official certifies that the information provided herein is true, accurate, and complete to the best of his knowledge.

Date: May 24, 1999

arome H. Heckman

Counsel for Ticona

14. Appendix

Attached are data on extraction of norbornene from ethylene-norbornene copolymer test specimens to aqueous media. (These data are referenced in footnote 2, above.)

Final Report

Determination of the Potential Migration of Norbornene and Decalin from Cyclic Olefin Copolymers into Food-Simulating Solvents

PREPARED FOR: Ticona

COVANCE STUDY NUMBER: 6954-101



Sponsor:

Ticona Summit, New Jersey Covance Laboratories Inc.
P.O. Box 7545
Madison, Wisconsin 53707-7545
Packages: 3301 Kinsman Boulev

Madison, Wisconsin 53704

Tel: 608/241-4471 Fax: 608/241-7227

FINAL REPORT

Study Title:

Determination of the Potential Migration of Norbornene and Decalin from Cyclic Olefin Copolymers into Food-Simulating Solvents

Authors:

Jessie L. Nelson Melanie M. McCort-Tipton

Study Completion Date:

May 21, 1999

Performing Laboratory:

Covance Laboratories Inc. 3301 Kinsman Boulevard Madison, Wisconsin 53704

Laboratory Project Identification

Covance 6954-101

Page 1 of 48

000315

THE AMERICAS EUROPE ASIA/PACIFIC AFRICA

STUDY IDENTIFICATION

Determination of the Potential Migration of Norbornene and Decalin from Cyclic Olefin Copolymers into Food-Simulating Solvents

Test Articles

Topas® cyclic olefin polymers (COC)

Sponsor

Ticona

86 Morris Avenue

Summit, New Jersey 07901

Study Monitor

Gerald S. Kirshenbaum

Ticona

86 Morris Avenue

Summit, New Jersey 07901

Telephone Number: (908) 522-7662 Facsimile Number: (908) 522-3932

Study Director

Melanie M. McCort-Tipton

Covance Laboratories Inc.

P.O. Box 7545

Madison, Wisconsin 53707-7545 Telephone Number: (608) 664-3049 Facsimile Number: (608) 664-3022

Study Locations

Covance Laboratories Inc. 3301 Kinsman Boulevard Madison, Wisconsin 53704

Covance Laboratories Inc.

802 Deming Way

Madison, Wisconsin 53717

Study Timetable

Experimental Start Date

Experimental Termination Date

December 15, 1998

February 8, 1999

KEY PERSONNEL

Agrosciences and Industrial Chemistry

Keith R. Rowley Associate Director

Melanie M. McCort-Tipton Staff Scientist/Manager

Paul Severin Research Assistant

Carol Mesun Study Coordinator

Jessie Nelson Study Coordinator

CONTENTS

	Page
STUDY IDENTIFICATION	2
KEY PERSONNEL	3
CONTENTS	4
ABSTRACT	6
INTRODUCTION	8
REGULATORY COMPLIANCE	8
MATERIALS	8
Test Articles Test Article Identification Analytical Standard Test System Extraction Cells	8 8 9 9
PROCEDURE	9
Phase I - Method Development Phase II - Extraction/Analysis Phase III - Validation	9 9 10
RESULTS	10
Phase I - Method Development Phase II - Extraction/Analysis Phase III - Validation	10 11 11
DATA AND SPECIMEN RETENTION	12
STATISTICAL EVALUATION	12
Table 1 Analysis of Norbornene in 10% Ethanol Extracts of Topas [®] 6015 Table 2 Analysis of Norbornene in 95% Ethanol Extracts of Topas [®] 6015 Table 3 Analysis of Norbornene in 10% Ethanol Extracts of Topas [®] 8007	13 14
Table 4 Analysis of Norbornene in 95% Ethanol Extracts of Topas 8007	15 16
Table 5 Analysis of Decalin in 10% Ethanol Extracts of Topas® 6015	17
Table 6 Analysis of Decalin in 95% Ethanol Extracts of Topas® 6015	18
Table 7 Analysis of Decalin in 10% Ethanol Extracts of Topas® 8007	19
Table 8 Analysis of Decalin in 95% Ethanol Extracts of Topas® 8007	20
Table 9 Validation of Norbornene in 10% Ethanol Extracts of Topas® 6015	21
Table 10 Validation of Norbornene in 95% Ethanol Extracts of Topas® 6015	22

	Covance 6954-101
Table 11 Validation of Norbornene in 10% Ethanol Extracts of Topas® 8007	23
Table 12 Validation of Norbornene in 95% Ethanol Extracts of Topas [®] 8007	24
Table 13 Validation of Decalin in 10% Ethanol Extracts of Topas [®] 6015	25
Table 14 Validation of Decalin in 95% Ethanol Extracts of Topas [®] 6015	26
Table 15 Validation of Decalin in 10% Ethanol Extracts of Topas [®] 8007	27
Table 16 Validation of Decalin in 95% Ethanol Extracts of Topas® 8007	28
SIGNATURES	29
APPENDIX A Methods	30
APPENDIX B. Example Standard Curve and Chromatography	33

ABSTRACT

This report describes a study that was performed on Topas® cyclic olefin copolymers (COC). The purpose of this study was to determine the potential migration of norbornene and decalin from cyclic olefin copolymers into food-simulating solvents under exaggerated conditions of use.

Analytical methodology was developed to determine norbornene and decalin in 10% and 95% ethanol extracts. The limit of detection (LOD) was 50 ppb $(0.5 \,\mu\text{g/in.}^2)$ for each analyte.

The test articles were extracted, in triplicate, under the following conditions.

Test Article	Solvent	Temperature/Time
Topas® 6015	10% Ethanol	121°C for 2 hours, then 40°C for 238 Hours
	95% Ethanol	121°C for 2 hours, then 40°C for 238 Hours
Topas® 8007	10% Ethanol	66°C for 2 hours, then 40°C for 238 Hours
	95% Ethanol	66°C for 2 hours, then 40°C for 238 Hours

Samples were removed after 2-, 24-, 96-, and 240-hours of exposure and analyzed for norbornene and decalin. The mean concentrations are presented below and on the following page.

Sample	Covance	Norbornene Mean Concentration (µg/in. ²)					
Identification	Log Number	2 Hours	24 Hours	96 Hours	240 Hours		
10% Ethanol							
Topas 6015	8-7972	< 0.501	< 0.501	<0.501	<0.501		
Topas 8007	8-7973	<0.501	<0.501	<0.501	<0.501		
		95% Etha	anol		₹		
Topas 6015	8-7972	< 0.501	< 0.501	<0.501	<0.501		
Topas 8007	8-7973	< 0.501	<0.501	<0.501	<0.501		

Sample	Covance	Decalin Mean Concentration (µg/in.2)					
Identification	Log Number	2 Hours	24 Hours	96 Hours	240 Hours		
10% Ethanol							
Topas 6015	8-7972	<0.501	<0.501	<0.501	<0.501		
Topas 8007	8-7973	<0.501	<0.501	<0.501	<0.501		
		95% Etha	anol				
Topas 6015	8-7972	<0.501	<0.501	<0.501	<0.501		
Topas 8007	8-7973	< 0.501	< 0.501	< 0.501	< 0.501		

Since the analytes were not detected in the extracts, validations were conducted by spiking the 240-hour extracts with norbornene and decalin at the LOD. All fortifications contained detectable levels of the analytes.

INTRODUCTION

This report describes a study that was performed on Topas® cyclic olefin copolymers (COC). The purpose of this study was to determine the potential migration of norbornene and decalin from cyclic olefin copolymers into food-simulating solvents under exaggerated conditions of use.

The study was divided into three phases.

- Phase I Method Development
- Phase II Extraction/Analysis
- Phase III Validation

REGULATORY COMPLIANCE

This study was conducted in accordance with the Food and Drug Administration (FDA) "Recommendations for Chemistry Data for Indirect Food Additive Petitions," (June 1995).

MATERIALS

Test Articles

The test articles consisted of Topas® cyclic olefin polymers, supplied by the Sponsor. A description of each polymer is presented below. The test articles were at least 20 mils thick.

Sample Identification	Description	Covance Log Number
Topas® 8007:	Uses 36% norbornene; HDT of 75°C	8-7972
Topas® 6015:	HDT of 150°C	8-7973

The test articles were stored under ambient conditions prior to testing. Information on the purity and stability of the test articles at ambient conditions is the responsibility of the Sponsor.

Test Article Identification

Each test article was identified by a unique Covance identification number. The log numbers are presented in the table above.

Analytical Standard

Analytical standards, of known purity, were obtained commercially for the following.

- Norbornene (norbornylene), 99%, Acros Organic Chemicals
- Decalin (decahydronaphthalene), 98%, Acros Organic Chemicals

Test System

Food Simulating Solvents:

- Ethanol, 95%, Aaper Alcohol Company, Shelbyville, Kentucky
- Ethanol, 10%, prepared from 95% ethanol and water processed through a Milli-Q purification system, Millipore Corporation, Bedford, Massachusetts

Extraction Cells

The two-sided extraction cells consisted of appropriate enclosures capable of withstanding the high pressures generated by heating the extraction solvents past their boiling points. Two sides of the test articles, 120 in.², were exposed to 700 mL of extraction solvent.

Safety Precautions

Adequate procedures were taken to ensure worker safety and were based on information contained in Material Safety Data Sheets supplied by the Sponsor, as well as those data sheets for the solvents themselves.

PROCEDURE

Phase I - Method Development

Analytical methodology was developed to determine norbornene and decalin in 10% and 95% ethanol extracts. The limit of detection (LOD) was 50 ppb (0.5 µg/in.²). A preliminary validation of the methods and the linearity of the standard response versus concentration was established.

Phase II - Extraction/Analysis

The test articles were extracted, in triplicate, under the conditions outlined on the following page.

Test Article	Solvent	Temperature/Time
Topas® 6015	10% Ethanol	121°C for 2 hours, then 40°C for 238 Hours
	95% Ethanol	121°C for 2 hours, then 40°C for 238 Hours
Topas® 8007	10% Ethanol	66°C for 2 hours, then 40°C for 238 Hours
	95% Ethanol	66°C for 2 hours, then 40°C for 238 Hours

Samples were removed after 2-, 24-, 96-, and 240-hours of exposure and analyzed for norbornene and decalin using the method developed in Phase I. Each replicate contained sufficient test article to detect the appropriate residue level. The ratio of solvent to surface area was 5.8 mL of solvent per square inch of test article exposed.

Phase III - Validation

Validations were conducted by spiking the 240-hour extracts, in triplicate, with known concentrations of analytes at approximately one half, one, and two times the amount detected. If the analyte was not detected, then the validations were conducted, in triplicate, at the LOD.

RESULTS

Phase I - Method Development

Methods were developed to determine norbornene and decalin in 10% and 95% ethanol extracts. These methods are presented in Appendix A. Example standard curves demonstrating the linearity of the standards response versus concentration are presented in Appendix B.

Phase II - Extraction/Analysis

The mean concentrations detected in the extracts are presented below. The individual results are presented in Tables 1-4 for norbornene and Tables 5-8 for decalin. Example chromatography is presented in Appendix B.

Sample	Covance	Norbornene Mean Concentration (µg/in. ²)					
Identification	Log Number	2 Hours	24 Hours	96 Hours	240 Hours		
10% Ethanol							
Topas 6015	8-7972	<0.501	<0.501	<0.501	<0.501		
Topas 8007	8-7973	<0.501	<0.501	<0.501	<0.501		
		95% Ethan	<u>nol</u>				
Topas 6015	8-7972	<0.501	<0.501	<0.501	<0.501		
Topas 8007	8-7973	<0.501	<0.501	<0.501	<0.501		
	Covance Decalin Mean Concentration (µg/in.²)						
Sample	Covance	Decali	n Mean Conc	entration (µ)	2/in. ²)		
Sample Identification	Covance Log Number	Decali 2 Hours	n Mean Conc 24 Hours	entration (με 96 Hours	g/in. ²) 240 Hours		
•	_		24 Hours				
•	_	2 Hours	24 Hours				
Identification	Log Number	2 Hours 10% Ethan	24 Hours	96 Hours	240 Hours		
Identification Topas 6015	Log Number 8-7972	2 Hours 10% Ethan <0.501	24 Hours nol <0.501 <0.501	96 Hours <0.501	<0.501 <0.501		
Identification Topas 6015	Log Number 8-7972	2 Hours 10% Ethan <0.501 <0.501	24 Hours nol <0.501 <0.501	96 Hours <0.501	240 Hours <0.501		

Phase III - Validation

Since the analytes were not detected in the extracts, validations were conducted by spiking the 240-hour extracts with norbornene and decalin at the LOD. All fortifications contained

detectable levels of the compounds. The individual results are presented in Tables 9-16, while example chromatography is presented in Appendix B.

DATA AND SPECIMEN RETENTION

When the final report is completed, the items to be transferred to and maintained in the archives of Covance will include, but will not be limited to:

- Protocol
- Test article information
- Test article preparation records
- Test article analysis records
- Study correspondence
- Final report

The following supporting records to be retained at Covance but not archived with the study data will include, but not be limited to:

- Storage location temperature records
- Instrument calibration and maintenance records

Upon acceptance of the final report, any remaining test article will be returned to the Sponsor, or at the Sponsor's request, destroyed.

All raw data, documentation, records, protocols, and final reports generated as a result of this study will be archived by Covance for a period of one year following signing of the final report. At least one year after signing of the final report, all of the aforementioned materials, with the exception of the original final report, protocol, amendments (if necessary), and correspondence, will be sent to a place designated by the Sponsor and a fee will be charged. The Sponsor may elect to have the materials retained in the Covance archives for an additional period of time and Covance will charge a storage fee. If the Sponsor chooses to have Covance dispose of the materials, a disposal fee will be charged.

STATISTICAL EVALUATION

The mean, standard deviation, and relative standard deviation will be calculated for each group of at least three analyses, where applicable.

Table 1

Analysis of Norbornene in 10% Ethanol Extracts of Topas® 6015

Sample	Covance		Concer	ntration	Mean		
Identification	Log Number	Replicate	(μg/mL)	$(\mu g/in.^2)^a$	$(\mu g/in.^2)$		
2 Hours at 121°C							
Topas 6015	8-7972	1	< 0.0858	< 0.501	<0.501		
Lot No.970320		2	< 0.0858	< 0.501			
		3	< 0.0858	< 0.501			
Solvent	NA	1	< 0.0858				
Blank		2	< 0.0858				
	2 Hours at	121°C, then	22 hours at 40	<u> </u>			
Topas 6015	8-7972	1	< 0.0858	< 0.501	<0.501		
Lot No.970320		2	<0.0858	< 0.501			
		3	< 0.0858	< 0.501			
Solvent	NA	1	< 0.0858				
Blank		2	< 0.0858				
	2 Hours at	121°C, then	94 hours at 40	<u>°C</u>			
Topas 6015	8-7972	1	< 0.0858	< 0.501	< 0.501		
Lot No.970320		2	< 0.0858	< 0.501			
		3	<0.0858	< 0.501			
Solvent	NA	1	< 0.0858				
Blank		2	< 0.0858				
	2 Hours at	121°C, then 2	38 hours at 40	<u>0°C</u>			
Topas 6015	8-7972	1	<0.0858	< 0.501	<0.501		
Lot No.970320		2	< 0.0858	< 0.501			
		3	< 0.0858	< 0.501			
Solvent	NA	1	< 0.0858				
Blank		2	< 0.0858				

a Concentration ($\mu g/\text{in.}^2$) = Concentration ($\mu g/\text{mL}$) x (700 mL/120 in.²)

Table 2

Analysis of Norbornene in 95% Ethanol Extracts of Topas® 6015

Sample	Covance		Concer	Mean	
Identification	Log Number	Replicate	$(\mu g/mL)$	$(\mu g/in.^2)^a$	$\mu g/in.^2$
		2 Hours at 1	21°C		
Topas 6015	8-7972	1	<0.0858	< 0.501	< 0.501
Lot No.970320		2	< 0.0858	< 0.501	
		3	<0.0858	< 0.501	
Solvent	NA	1	<0.0858		
Blank		2	<0.0858		
	2 Hours at	: 121°C, then 2	22 hours at 40	°C	
Topas 6015	8-7972	1	<0.0050		<0.501
Lot No.970320	0-1912		<0.0858 <0.0858	< 0.501	<0.501
LUI NO.970320		2 3	<0.0858	< 0.501	
		٦	~0.0636	< 0.501	
Solvent	NA	1	< 0.0858		
Blank		2	<0.0858		
	2 Hours at	121°C, then	94 hours at 40	<u>°C</u>	
Topas 6015	8-7972	1	<0.0858	< 0.501	<0.501
Lot No.970320		2	< 0.0858	< 0.501	0.501
		3	<0.0858	< 0.501	
Solvent	NA	1	<0.0858		
Blank	1471	2	<0.0858		
Dium					
	2 Hours at	121°C, then 2	38 hours at 40	<u>0°C</u>	
Topas 6015	8-7972	1	<0.0858	< 0.501	<0.501
Lot No.970320		2	<0.0858	< 0.501	
		3	<0.0858	< 0.501	
Solvent	NA	Ī	< 0.0858		
Blank		2	<0.0858		

a Concentration ($\mu g/in.^2$) = Concentration ($\mu g/mL$) x (700 mL/120 in.²)

Table 3

Analysis of Norbornene in 10% Ethanol Extracts of Topas® 8007

Sample	Covance		Concer	ntration	Mean
Identification	Log Number	Replicate	(μg/mL)	$(\mu g/in.^2)^a$	$(\mu g/in.^2)$
		2 Hours at 6	6°C		
Topas 8007	8-7973	1	< 0.0858	< 0.501	< 0.501
Lot No. 970205		2	<0.0858	< 0.501	10.501
		3	<0.0858	< 0.501	
Solvent	NA	1	<0.0858		
Blank		2	<0.0858		
	2 Hours at	t 66°C, then 2	2 hours at 40°	<u>C</u>	
Topas 8007	8-7973	1	<0.0858	<0.501	<0.501
Lot No. 970205		2	< 0.0858	< 0.501	\0.501
		3	<0.0858	< 0.501	
Solvent	NA	1	< 0.0858		
Blank		2	< 0.0858		
	2 Hours at	66°C, then 9	4 hours at 40°	<u>C</u>	
Topas 8007	8-7973	1	<0.0858	<0.501	< 0.501
Lot No. 970205		2	< 0.0858	< 0.501	7,5 0 2
		3	<0.0858	< 0.501	
Solvent	NA	1	<0.0858		
Blank		2	<0.0858		
	2 Hours at	66°C, then 23	8 hours at 40	<u>°C</u>	
Topas 8007	8-7973	1	<0.0858	<0.501	<0.501
Lot No. 970205		2	< 0.0858	< 0.501	
		3	<0.0858	< 0.501	
Solvent	NA	1	<0.0858		
Blank		2	<0.0858		

a Concentration ($\mu g/in.^2$) = Concentration ($\mu g/mL$) x (700 mL/120 in.²)

Table 4

Analysis of Norbornene in 95% Ethanol Extracts of Topas® 8007

Covance		Concer	ntration	Mean
Log Number	Replicate	(μg/mL)	$(\mu g/in.^2)^a$	$(\mu g/in.^2)$
	2 Hours at 6	<u>6°C</u>		
8-7973	1	<0.0858	< 0.501	<0.501
	2	< 0.0858		0.501
	3	<0.0858	< 0.501	
NA	1	<0.0858		
	2	<0.0858		
2 Hours at	t 66°C, then 2	2 hours at 40°	<u>C</u>	
8-7973	1	< 0.0858	<0.501	<0.501
				-0.501
	3	<0.0858	< 0.501	
NA	1	<0.0858		
	2	< 0.0858		
2 Hours at	t 66°C, then 9	4-hours at 40°	\mathbf{c}	
8.7072	1	<0.0050	<0.501	<0.501
0-1313				< 0.501
	3			
37.4			0.001	
NA				
	2	<0.0858		
2 Hours at	66°C, then 23	8 hours at 40	<u>°C</u>	
8-7973	1	<0.0858	< 0.501	<0.501
	2	< 0.0858	< 0.501	
	3	<0.0858	< 0.501	
NA	1	<0.0858		
	2	<0.0858		
	8-7973 NA 2 Hours at 8-7973 NA 2 Hours at 8-7973 NA 2 Hours at 8-7973	Log Number Replicate 2 Hours at 6 8-7973 1 2 3 NA 1 2 8-7973 1 2 2 Hours at 66°C, then 9 8-7973 1 2 3 NA 1 2 2 Hours at 66°C, then 9 8-7973 1 2 2 Hours at 66°C, then 23 8-7973 1 2 3 NA 1 2 3 NA 1	Log Number Replicate (μg/mL)	Log Number Replicate (μg/mL) (μg/in.²)* 2 Hours at 66°C 8-7973 1 <0.0858

a Concentration ($\mu g/in.^2$) = Concentration ($\mu g/mL$) x (700 mL/120 in.²)

Table 5

Analysis of Decalin in 10% Ethanol Extracts of Topas® 6015

Sample	Covance		Conce	ntration	Mean				
Identification	Log Number	Replicate	(μg/mL)	$(\mu g/in.^2)^a$	$(\mu g/in.^2)$				
		2 Hours at	121°C						
Topas 6015	8-7972	1	<0.0858	< 0.501	< 0.501				
Lot No.970320		2	< 0.0858	< 0.501					
		3	<0.0858	< 0.501					
Solvent	NA	1	<0.0858						
Blank		2	<0.0858						
	2 Hours	at 121°C, ther	22 hours at 4	<u>0°C</u>					
Topas 6015	8-7972	1	<0.0858	< 0.501	< 0.501				
Lot No.970320		2	< 0.0858	< 0.501					
		3	<0.0858	< 0.501					
Solvent	NA	1	<0.0858						
Blank		2	<0.0858						
	2 Hours	at 121°C, ther	94 hours at 4	<u>0°C</u>					
Topas 6015	8-7972	1	<0.0858	<0.501	< 0.501				
Lot No.970320		2	< 0.0858	< 0.501					
		3 .	<0.0858	< 0.501					
Solvent	NA	1	<0.0858						
Blank		2	<0.0858						
2 Hours at 121°C, then 238 hours at 40°C									
Topas 6015	8-7972	1	<0.0858	< 0.501	< 0.501				
Lot No.970320		2	< 0.0858	< 0.501					
		3	< 0.0858	< 0.501					
Solvent	NA	1	<0.0858						
Blank		2	< 0.0858						

a Concentration ($\mu g/in.^2$) = Concentration ($\mu g/mL$) x (700 mL/120 in.²)

Table 6

Analysis of Decalin in 95% Ethanol Extracts of Topas® 6015

Sample	Covance		Conce	ntration	Mean
Identification	Log Number	Replicate	$\mu g/mL$	$(\mu g/in.^2)^a$	$\mu g/in.^2$
Topas 6015	8-7972	1	< 0.0858	< 0.501	< 0.501
Lot No.970320		2	< 0.0858	< 0.501	
		3	< 0.0858	< 0.501	
Solvent	NA	1	< 0.0858		
Blank		2	<0.0858		
	2 Hours	at 121°C, ther	n 22 hours at 4	<u>0°C</u>	
Topas 6015	8-7972	1	<0.0858	<0.501	<0.501
Lot No.970320		2	< 0.0858	< 0.501	*****
		3	<0.0858	< 0.501	
Solvent	NA	1	<0.0858		
Blank		2	<0.0858		
	2 Hours	at 121°C, ther	1 94 hours at 4	<u>0°C</u>	
Topas 6015	8-7972	1	<0.0858	< 0.501	<0.501
Lot No.970320		2	< 0.0858	< 0.501	0.501
		3	< 0.0858	< 0.501	
Solvent	NA	1	<0.0858		
Blank		2	<0.0858		
	2 Hours a	t 121°C, then	238 hours at 4	10°C	
Topas 6015	8-7972	1	<0.0858	< 0.501	<0.501
Lot No.970320		2	<0.0858	< 0.501	-0.501
		3	< 0.0858	< 0.501	
Solvent	NA	1	< 0.0858		
Blank	- ·	2	< 0.0858		

a Concentration ($\mu g/in.^2$) = Concentration ($\mu g/mL$) x (700 mL/120 in.²)

Table 7

Analysis of Decalin in 10% Ethanol Extracts of Topas® 8007

Sample	Covance		Concer	ntration	Mean
Identification	Log Number	Replicate	(μg/mL)	$(\mu g/in.^2)^a$	μ g/in. ²)
		2 Hours at	<u>66°C</u>		***************************************
Topas 8007	8-7973	1	< 0.0858	< 0.501	< 0.501
Lot No. 970205		2	< 0.0858	< 0.501	
		3	<0.0858	< 0.501	
Solvent	NA	1	<0.0858		
Blank		2	<0.0858		
	2 Hours	at 66°C, then	22 hours at 40	<u>)°C</u>	
Topas 8007	8-7973	1	<0.0858	< 0.501	<0.501
Lot No. 970205		2	< 0.0858	< 0.501	3.3 3 1
		3	<0.0858	< 0.501	
Solvent	NA	1	<0.0858		
Blank		2	<0.0858		
	2 Hours	at 66°C, then	94 hours at 40	<u>)°C</u>	
Topas 8007	8-7973	1	<0.0858	< 0.501	<0.501
Lot No. 970205		2	< 0.0858	< 0.501	
		3	<0.0858	< 0.501	
Solvent	NA	1	<0.0858		
Blank		2	<0.0858		
	2 Hours	at 66°C, then	238 hours at 4	<u>0°C</u>	
Topas 8007	8-7973	1	<0:0858	< 0.501	< 0.501
Lot No. 970205		2	< 0.0858	< 0.501	3.531
		3	<0.0858	< 0.501	
Solvent	NA	1	<0.0858		
Blank	- · - -	2	< 0.0858		

a Concentration ($\mu g/in.^2$) = Concentration ($\mu g/mL$) x (700 mL/120 in.²)

Table 8

Analysis of Decalin in 95% Ethanol Extracts of Topas® 8007

Sample	Covance		Conce	ntration	Mean			
Identification	Log Number	Replicate	$(\mu g/mL)$	$(\mu g/in.^2)^a$	$(\mu g/in.^2)$			
		2 Hours at	: 66°C					
Topas 8007	8-7973	1	< 0.0858	< 0.501	< 0.501			
Lot No. 970205		2	< 0.0858	< 0.501				
		3	<0.0858	< 0.501				
Solvent	NA	1	<0.0858					
Blank		2	<0.0858					
	2 Hours	at 66°C, then	22 hours at 40	<u>)°C</u>				
Topas 8007	8-7973	1	<0.0858	<0.501	< 0.501			
Lot No. 970205		2	< 0.0858	< 0.501	3.532			
		3	<0.0858	< 0.501				
Solvent	NA	1	<0.0858					
Blank		2	< 0.0858					
	2 Hours	at 66°C, then	94 hours at 40	<u> </u>				
Topas 8007	8-7973	1	<0.0858	< 0.501	<0.501			
Lot No. 970205		2	< 0.0858	< 0.501				
		3	<0.0858	< 0.501				
Solvent	NA	1	<0.0858					
Blank		2	< 0.0858					
2 Hours at 66°C, then 238 hours at 40°C								
Topas 8007	8-7973	1	<0.0858	< 0.501	< 0.501			
Lot No. 970205		2	< 0.0858	< 0.501	0.501			
		3	<0.0858	< 0.501				
Solvent	NA	1	<0.0858					
Blank		2	< 0.0858					

a Concentration ($\mu g/in.^2$) = Concentration ($\mu g/mL$) x (700 mL/120 in.²)

Table 9

Validation of Norbornene in 10% Ethanol Extracts of Topas® 6015

Sample	Covance		Concentration Detect	Corrected Concentration ed Detected	Concentration Added	Percent	Mean
Identification	Log Number	Replicate	(μg/mL) (μg/in. ²	$\frac{(\mu g/in.^2)^b}{}$	(μg/in. ²)	Recovery	(%)
Topas 6015	8-7972	1	0.0570 c 0.333	0.333	0.501	Detected	Detected
Lot No.970320		2	0.0291 c 0.170	0.170	0.501	Detected	
LOD Spike		3	0.0858 c 0.501	0.501	0.501	Detected	
Topas 6015	8-7972	1	<0.0858 <0.503	l			
Lot No.970320		2	<0.0858 <0.503	İ			

a Concentration ($\mu g/\text{in.}^2$) = Concentration ($\mu g/\text{mL}$) x (700 mL/120 in.²)

b Concentration corrected by subtracting the mean level detected in the control, when necessary.

c The value was extrapolated below the standard curve.

22

Covance 6954-10

Table 10

Validation of Norbornene in 95% Ethanol Extracts of Topas® 6015

Sample	Covance		Concentration	on Detected	Corrected Concentration Detected	Concentration Added	Percent	Mean
Identification	Log Number	Replicate	(µg/mL)	$(\mu g/in.^2)^a$	$(\mu g/in.^2)^b$	$(\mu g/in.^2)$	Recovery	(%)
Topas 6015	8-7972	1	0.114	0.665	0.665	0.501	Detected	Detected
Lot No.970320		2	0.0951	0.555	0.555	0.501	Detected	
LOD Spike		3	0.0857 c	0.500	0.500	0.501	Detected	
Topas 6015	8-7972	1	<0.0858	<0.501				
Lot No.970320		2	<0.0858	< 0.501				

a Concentration ($\mu g/\text{in.}^2$) = Concentration ($\mu g/\text{mL}$) x (700 mL/1/20 in.²)

b Concentration corrected by subtracting the mean level detected in the control, when necessary.

c The value was extrapolated below the standard curve.

Table 11

Validation of Norbornene in 10% Ethanol Extracts of Topas® 8007

Sample	Covance		Concentration	n Detected	Corrected Concentration Detected	Concentration Added	Percent	Mean
Identification	Log Number	Replicate	(μg/mL)	$(\mu g/in.^2)^a$	$(\mu g/in.^2)^b$	(μg/in. ²)	Recovery	(%)
Topas 8007	8-7973	l	0.0699 с	0.408	0.408	0.501	Detected	Detected
Lot No. 970205		2	0.0829 c	0.484	0.484	0.501	Detected	
LOD Spike		3	0.0679 c	0.396	0.396	0.501	Detected	
Topas 8007	8-7973	1	<0.0858	<0.501				
Lot No. 970205		2	<0.0858	< 0.501				

a Concentration (μ g/in.²) = Concentration (μ g/mL) x (700 mL/120 in.²)

b Concentration corrected by subtracting the mean level detected in the control, when necessary.

c The value was extrapolated below the standard curve.

Table 12

Validation of Norbornene in 95% Ethanol Extracts of Topas® 8007

Sample	Covance		Concentration	on Detected	Corrected Concentration Detected	Concentration Added	Percent	Mean
Identification	Log Number	Replicate	(μg/mL)	(μg/in. ²) ^a	(μg/in. ²) ^b	(μg/in. ²)	Recovery	(%)
Topas 8007	8-7973	1	0.0792 с	0.462	0.462	0.501	Detected	Detected
Lot No. 970205		2	0.0685 c	0.400	0.400	0.501	Detected	
LOD Spike		3	0.101	0.589	0.589	0.501	Detected	
Topas 8007	8-7973	1	<0.0858	<0.501				
Lot No. 970205		2	<0.0858	< 0.501				

a Concentration ($\mu g/\text{in.}^2$) = Concentration ($\mu g/\text{mL}$) x (700 mL/120 in.²)

b Concentration corrected by subtracting the mean level detected in the control, when necessary.

c The value was extrapolated below the standard curve.

Table 13

Validation of Decalin in 10% Ethanol Extracts of Topas[®] 6015

Sample	Covance		Concentration	on Detected	Corrected Concentration Detected	Concentration Added	Percent	Mean
Identification	Log Number	Replicate	(μg/mL)	$(\mu g/in.^2)^a$	$(\mu g/in.^2)^b$	$\frac{(\mu g/in.^2)}{}$	Recovery	(%)
Topas 6015	8-7972	1	0.0626 c	0.365	0.365	0.501	Detected	Detected
Lot No.970320		2	0.0368 c	0.215	0.215	0.501	Detected	
LOD Spike		3	0.108	0.630	0.630	0.501	Detected	
Topas 6015	8-7972	1	<0.0858	<0.501				
Lot No.970320		2	< 0.0858	< 0.501				

a Concentration (μ g/in.²) = Concentration (μ g/mL) x (700 mL/120 in.²)

b Concentration corrected by subtracting the mean level detected in the control, when necessary.

c The value was extrapolated below the standard curve.

Covance 6954-10

Table 14

Validation of Decalin in 95% Ethanol Extracts of Topas® 6015

Sample	Covance		Concentrati	on Detected	Corrected Concentration Detected	Concentration Added	Percent	Mean
Identification	Log Number	Replicate	(μg/mL)	$(\mu g/in.^2)^a$	$(\mu g/in.^2)^b$	$(\mu g/in.^2)$	Recovery	(%)
Topas 6015	8-7972	1	0.0826	0.482	0.482	0.501	Detected	Detected
Lot No.970320		2	0.0887	0.517	0.517	0.501	Detected	
LOD Spike		3	0.0893	0.521	0.521	0.501	Detected	
Topas 6015	8-7972	1	<0.0858	< 0.501				
Lot No.970320		2	< 0.0858	< 0.501				

a Concentration ($\mu g/in.^2$) = Concentration ($\mu g/mL$) x (700 mL/120 in.²)

b Concentration corrected by subtracting the mean level detected in the control, when necessary.

c The value was extrapolated below the standard curve.

Sample	Covance		Concentra	tio	n Detected	Corrected Concentration Detected	Concentration Added	Percent	Mean
Identification	Log Number	Replicate	(μg/mL)		$(\mu g/in.^2)^a$	$(\mu g/in.^2)^b$	$\frac{(\mu g/in.^2)}{}$	Recovery	(%)
Topas 8007	8-7973	1	0.0731	c	0.426	0.426	0.501	Detected	Detected
Lot No. 970205		2	0.0912		0.532	0.532	0.501	Detected	
LOD Spike		3	0.0715	c	0.417	0.417	0.501	Detected	
Topas 8007	8-7973	1	< 0.0858		< 0.501				
Lot No. 970205		2	< 0.0858		< 0.501				

Table 15

Validation of Decalin in 10% Ethanol Extracts of Topas[®] 8007

a Concentration ($\mu g/in.^2$) = Concentration ($\mu g/mL$) x (700 mL/120 in.²)

b Concentration corrected by subtracting the mean level detected in the control, when necessary.

c The value was extrapolated below the standard curve.

Covance 6954-101

Table 16

Validation of Decalin in 95% Ethanol Extracts of Topas® 8007

Sample	Covance		Concentra	tior	1 Detected	Corrected Concentration Detected	Concentration Added	Percent	Mean
Identification	Log Number	Replicate	(µg/mL)		$(\mu g/in.^2)^a$	$\underline{(\mu g/in.^2)^b}$	$\frac{(\mu g/in.^2)}{}$	Recovery	(%)
Topas 8007	8-7973	1	0.0808	c	0.471	0.471	0.501	Detected	Detected
Lot No. 970205		2	0.0745	c	0.435	0.435	0.501	Detected	
LOD Spike		3	0.0716	С	0.418	0.418	0.501	Detected	
Topas 8007	8-7973	1	< 0.0858		< 0.501				
Lot No. 970205		2	< 0.0858		< 0.501				

a Concentration ($\mu g/in.^2$) = Concentration ($\mu g/mL$) x (700 mL/120 in.²)

000342

b Concentration corrected by subtracting the mean level detected in the control, when necessary.

c The value was extrapolated below the standard curve.

SIGNATURES

Melanie M. McCort-Tipton

Study Director

Covance Laboratories Inc.

5/2//99 Date

Keith R. Rowley

Associate Director/

Covance Laboratories Inc.

Date /

APPENDIX A

Methods

Analysis of Norbornene and Decalin in 10% Ethanol Extracts

A solid phase micro-extraction (SPME) fiber was used for analysis of norbornene and decalin in the 10% ethanol extracts. The SPME fiber is a fused silica fiber coated with 100 µm polydimethylsiloxane. Three milliliters of extract was placed in a 1-dram vial, containing a Teflon[®] coated stir bar, with a septum screw cap. The 1-dram vial was placed on a magnetic stir plate. The SPME needle was inserted through the septum into the stirring solution. The fiber remained immersed in the solution for 17 minutes, after which the analytes were thermally desorbed in the gas chromatograph (GC) injection port for 0.40 minutes. The GC parameters are presented below.

GC Parameters

Column DB-1 (30 m x 0.53 mm)

3 µm film thickness

Detector: Flame ionization

Temperatures:

Column: 65°C for 5 minutes, then 15°C/minute to

215°C, 215°C for 1 minute

Injector: 250°C Detector: 300°C

Flows:

Carrier: 8 mL/minute Helium
Makeup: 30 mL/minute Nitrogen

Air: 380 mL/minute Hydrogen: 37 mL/minute

Analysis of Norbornene and Decalin in 95% Ethanol Extracts

An aliquot of the 95% ethanol extract was placed in an autosampler vial and analyzed under the following GC parameters.

GC Parameters

Column

 $DB-1 (30 \text{ m} \times 0.53 \text{ mm})$

3 µm film thickness

Detector:

Flame ionization

Temperatures:

Column:

50°C for 5 minutes, then 10°C/minute to

250°C, 250°C for 1 minute

Injector:

250°C

Detector:

300°C

Flows:

Carrier:

8 mL/minute Helium

Makeup:

30 mL/minute Nitrogen

Air: Hydrogen: 380 mL/minute 37 mL/minute

Injection volume:

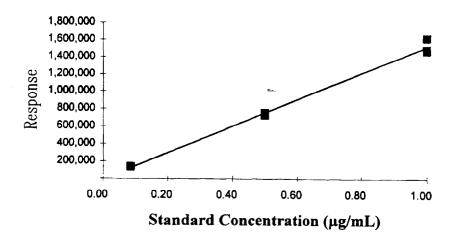
 $2 \mu L$

APPENDIX B

Example Standard Curve and Chromatography

Example Standard Curve for Norbornene in 10% Ethanol^a

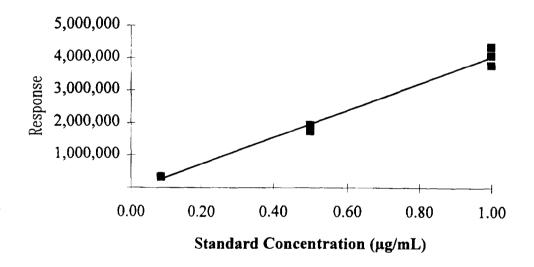
Standard Concentration	
$\mu g/mL$	Response
0.0858	131,805.5
0.0858	136,879.6
0.500	727,636.8
0.500	753,252.2
1.00	1,483,185.4
1.00	1,471,349.2
1.00	1,620,023.4



a This is a representative standard curve to demonstrate the linearity of the method.

Example Standard Curve for Decalin in 10% Ethanol^a

Response
_
319,118.1
315,755.6
1,745,313.8
1,941,128.0
4,082,608.6
3,779,481.5
4,349,379.3



Please note that the response is the sum of the cis and trans isomers of decalin.

a This is a representative standard curve to demonstrate the linearity of the method.

Figure B1 $0.0858~\mu\text{g/mL}~Norbornene~and~Decalin~Standard~in~10\%~Ethanol$

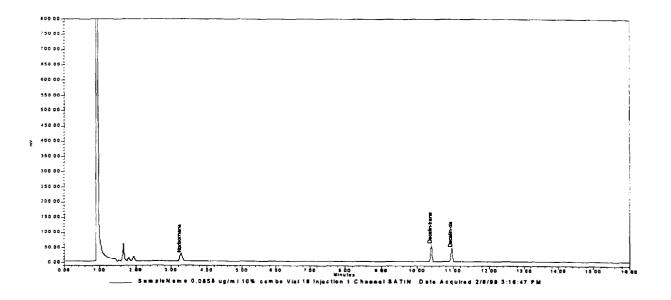


Figure B2 $0.500~\mu\text{g/mL}~Norbornene~and~Decalin~Standard~in~10\%~Ethanol$

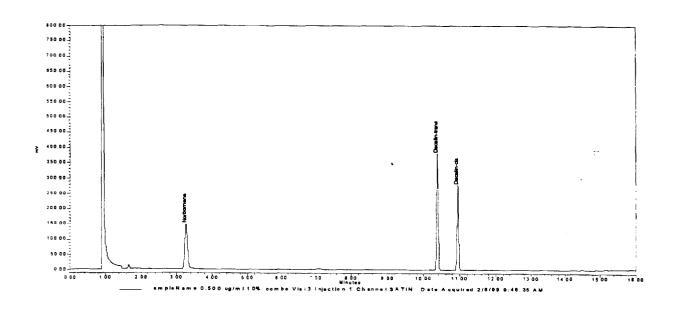


Figure B3 $1.00~\mu\text{g/mL}~Norbornene~and~Decalin~Standard~in~10\%~Ethanol$

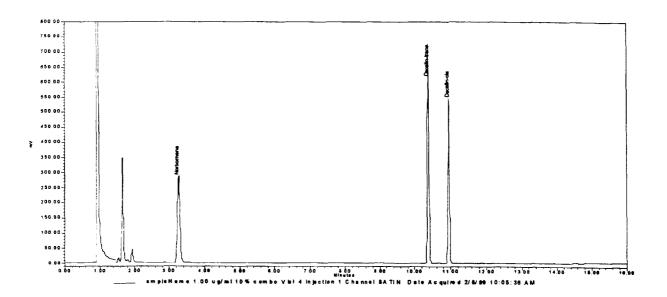


Figure B4

10% Ethanol Analysis, Solvent Blank, 240-Hour

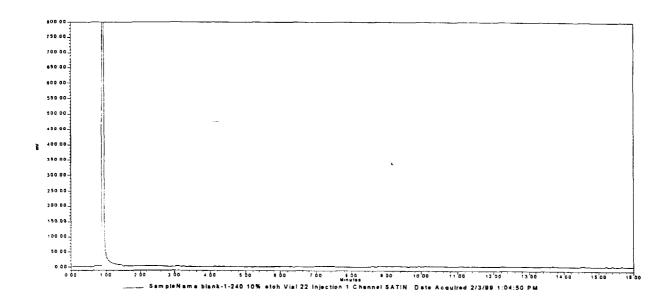


Figure B5

10% Ethanol Analysis, Topas® 6015, 8-7972, 240-Hour Extract

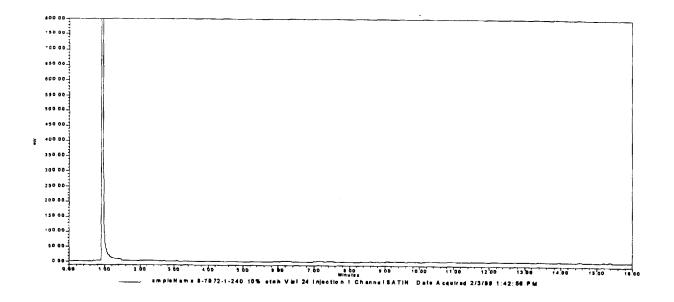


Figure B6
10% Ethanol Validation, Topas[®] 6015, 8-7972, 240-Hour Composite

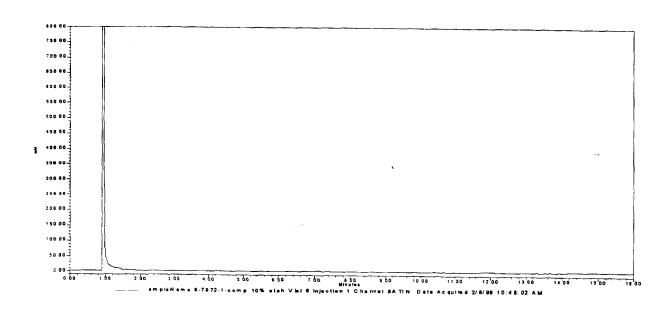


Figure B7

10% Ethanol Validation, Topas[©] 6015, 8-7972, LOD Spike

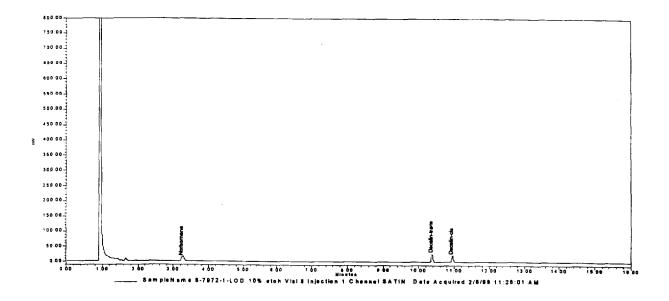


Figure B8

10% Ethanol Analysis, Topas® 8007, 8-7973, 240-Hour Extract

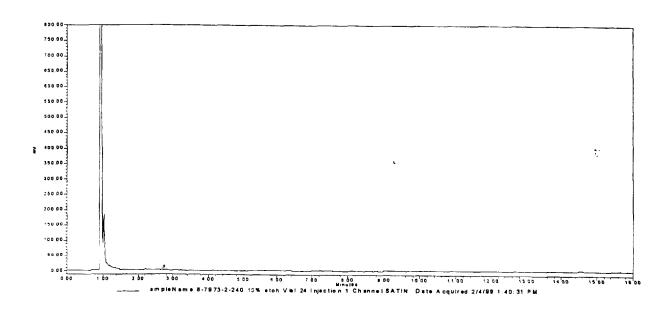


Figure B9

10% Ethanol Validation, Topas® 8007, 8-7973, 240-Hour Composite

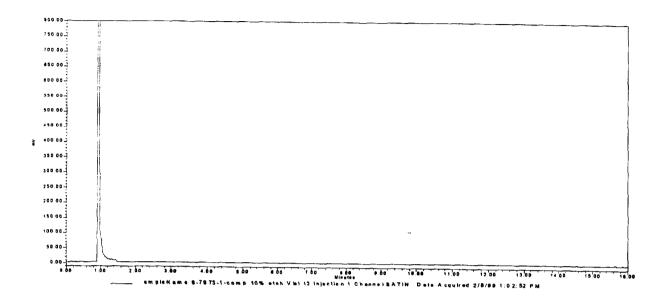
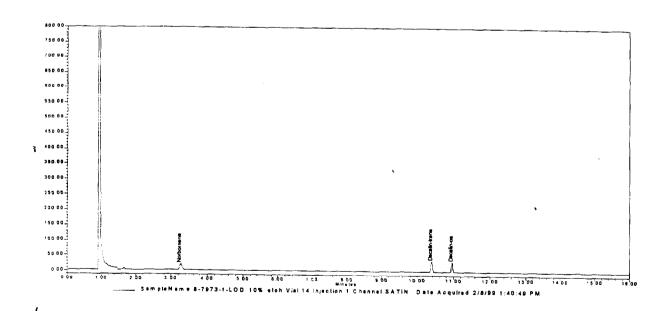


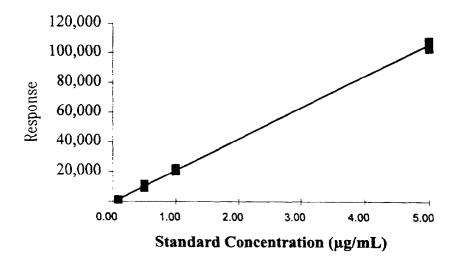
Figure B10

10% Ethanol Validation, Topas[©] 8007, 8-7973, LOD Spike



Example Standard Curve for Norbornene in 95% Ethanol^a

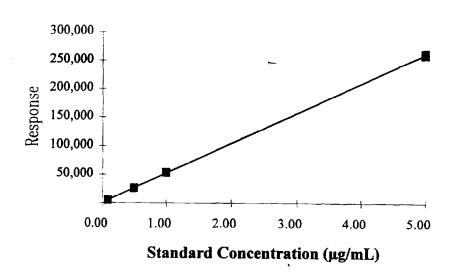
Standard Concentration (µg/mL)	Response
0.0858	1,081.7
0.0858	1,143.2
0.500	11,324.3
0.500	8,978.8
1.00	22,175.8
1.00	20,088.2
5.00	108,082.6
5.00	106,264.4
5.00	102,760.7



a This is a representative standard curve to demonstrate the linearity of the method.

Example Standard Curve for Decalin in 95% Ethanol^a

Standard Concentration	
(μg/mL)	Response
0.0858	4,982.7
0.0858	4,668.2
0.500	26,712.0
0.500	25,816.5
1.00	54,091.3
1.00	52,794.4
5.00	262,774.6
5.00	262,094.4
5.00	258,683.2



Please note that the response is the sum of the cis and trans isomers of decalin.

a This is a representative standard curve to demonstrate the linearity of the method.

ţ,

Figure B11 $0.0858~\mu\text{g/mL}~Norbornene~and~Decalin~Standard~in~95\%~Ethanol$

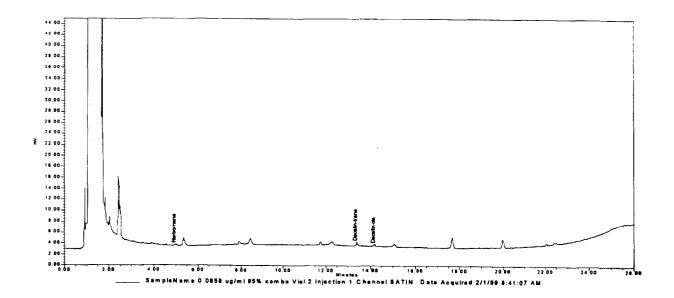


Figure B12 $0.500~\mu\text{g/mL}~Norbornene~and~Decalin~Standard~in~95\%~Ethanol$

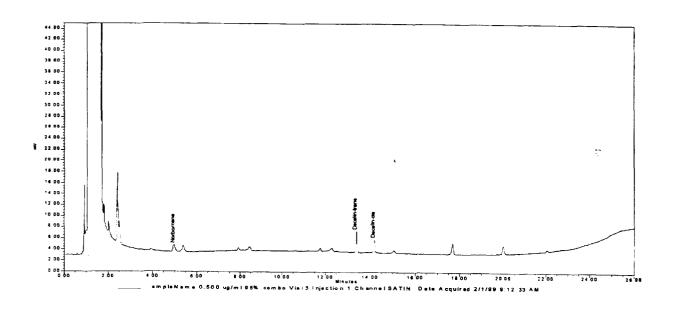


Figure B13 $1.00~\mu\text{g/mL Norbornene} \text{ and Decalin Standard in 95\% Ethanol}$

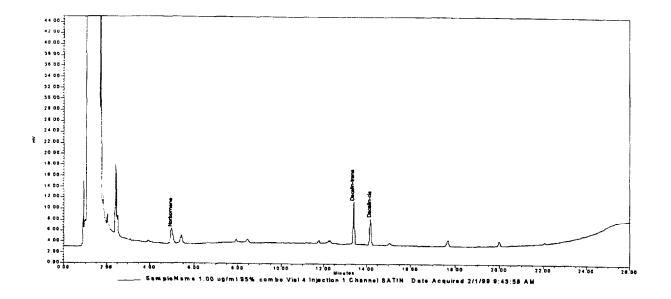


Figure B14 $$5.00\ \mu g/mL$ Norbornene and Decalin Standard in 95% Ethanol

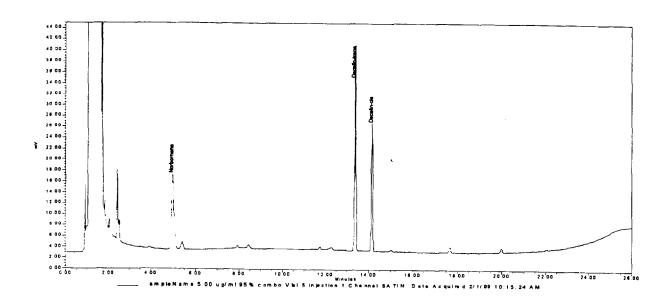


Figure B15
95% Ethanol Analysis, Solvent Blank, 240-Hour Extract

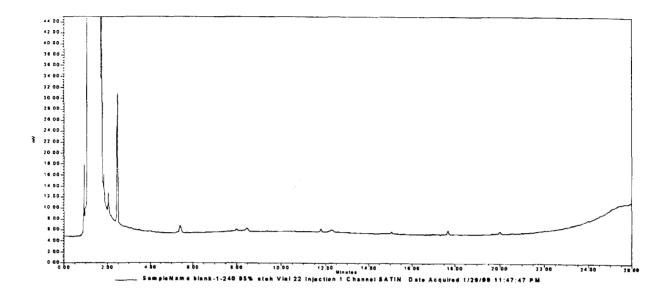


Figure B16

95% Ethanol Analysis, Topas® 6015, 8-7972, 240-Hour Extract

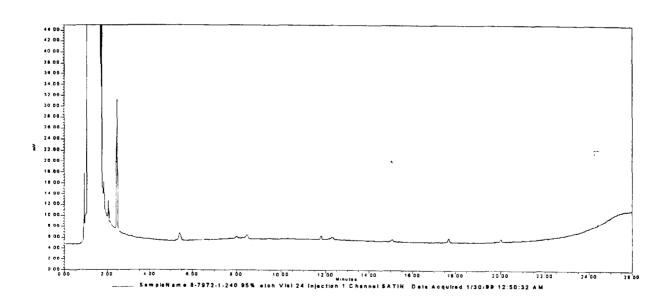


Figure B17
95% Ethanol Validation, Topas® 6015, 8-7972, 240-Hour Composite

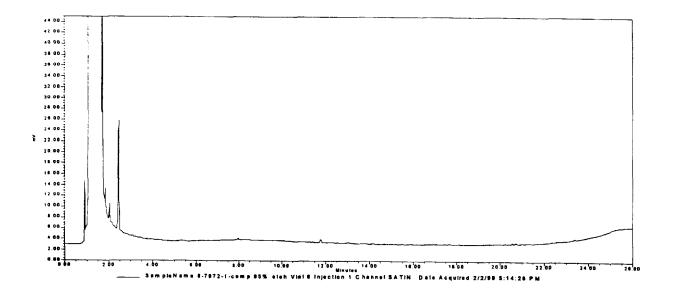


Figure B18

95% Ethanol Validation, Topas[©] 6015, 8-7972, LOD Spike

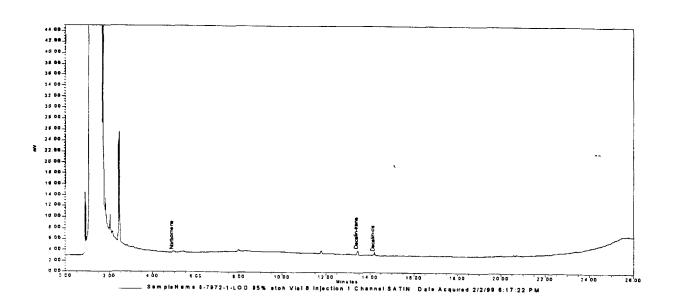


Figure B19
95% Ethanol Analysis, Topas® 8007, 8-7973, 240-Hour Extract

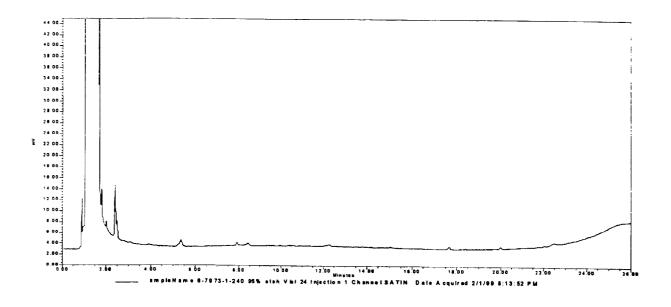


Figure B20

95% Ethanol Validation, Topas® 8007, 8-7973, 240-Hour Composite

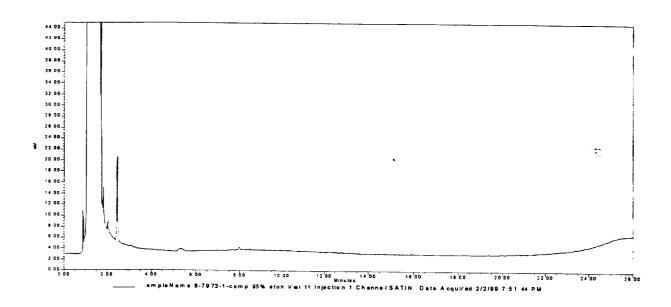


Figure B21
95% Ethanol Validation, Topas® 8007, 8-7973, LOD Spike

